



**Europäisches  
Patentamt**

**European  
Patent Office**

**Office européen  
des brevets**

20. 09. 2004

REC'D 23 NOV 2004

WIPO

PCT

**Bescheinigung**

**Certificate**

**Attestation**

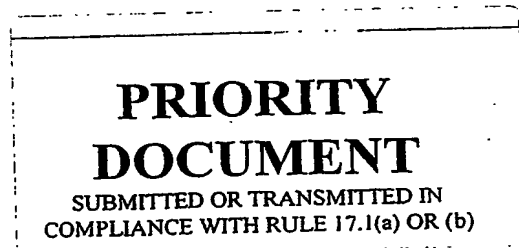
Die angehefteten Unterlagen stimmen mit der ursprünglich eingereichten Fassung der auf dem nächsten Blatt bezeichneten europäischen Patentanmeldung überein.

The attached documents are exact copies of the European patent application described on the following page, as originally filed.

Les documents fixés à cette attestation sont conformes à la version initialement déposée de la demande de brevet européen spécifiée à la page suivante.

**Patentanmeldung Nr. Patent application No. Demande de brevet n°**

03103472.1



Der Präsident des Europäischen Patentamts;  
Im Auftrag

For the President of the European Patent Office

Le Président de l'Office européen des brevets  
p.o.

**R C van Dijk**



Anmeldung Nr:  
Application no.: 03103472.1  
Demande no:

Anmeldetag:  
Date of filing: 19.09.03  
Date de dépôt:

Anmelder/Applicant(s)/Demandeur(s):

Tibotec Pharmaceuticals Ltd.  
Unit 4, Block 4B, Blanchardstown Corporate  
Park  
Blanchardstown, Dublin 15  
IRLANDE

Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:  
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.  
If no title is shown please refer to the description.  
Si aucun titre n'est indiqué se référer à la description.)

Multi-well assay for identifying the inhibition activity of compounds acting at  
different stages of microbial replication and having anti-microbial activity

In Anspruch genommene Priorität(en) / Priority(ies) claimed /Priorité(s)  
revendiquée(s)

Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/  
Classification internationale des brevets:

G01N33/48

Am Anmeldetag benannte Vertragstaaten/Contracting states designated at date of  
filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HU IE IT LU MC NL  
PT RO SE SI SK TR LI

**Multi-well assay for identifying the inhibition activity of compounds acting at different stages of microbial replication and having anti-microbial activity**

**5 Field of the invention**

The present invention relates to the medicinal field. In a first aspect, the present invention relates to an assay for identifying compounds inhibiting the replication cycle of a micro-organism e.g. AIDS. In another aspect, the invention relates to an apparatus for carrying out the assay according to the invention. The invention further  
10 relates to compounds identifiable with an assay according to the invention.

**Background**

A retrovirus, designated human immunodeficiency virus (HIV), is the etiological agent of the complex disease that includes progressive destruction of the immune system  
15 (acquired immune deficiency syndrome; AIDS) and degeneration of the central and peripheral nervous system. This virus is known by different names, including T-lymphocyte virus III (HTLV-III) or lymphadenopathy-associated virus (LAV) or AIDS-related virus (ARV) or human immunodeficiency virus (HIV). Up until now, two distinct families have been identified, i.e. HIV-1 and HIV-2. Hereinafter, HIV will be used to  
20 generically denote these viruses.

HIV is a type of retrovirus, belonging to single stranded RNA viruses, that can infect a number of different cells, including CD4 bearing macrophages and T-helper lymphocytes within a host. After initial contact and attachment to a cell of the immune  
25 system (e.g. lymphocytes, monocytes), there is a cascade of intracellular events. The end product of these events is the production of massive numbers of new viral particles, death of the infected cells, and ultimate devastation of the immune system.

The HIV replication cycle consists of different stages, which can be broadly identified  
30 as a) an entry stage, b) a reverse transcription stage, c) an integration stage, d) a budding stage and d) maturation (a protease activity stage).

In the entry stage, HIV enters a CD4 cell. This process as such also comprises different phases. In a first phase, a *CD4 receptor attachment phase*, HIV infection  
35 begins with the interaction of the HIV glycoprotein (gp) 120 with the CD4 receptor molecule on the surface of a target cell. In a second phase, a *co-receptor binding phase*, following CD4 binding, a center material change in the HIV gp 120/gp 41

complex is induced by interaction of gp 120 with the chemokine receptors CCR5 or CXCR4. This change in confirmation exposes gp 41 allowing it to initiate fusion of the membranes. In a third phase, a phase involving *membrane fusion events*, as the virus fuses with the cell, internalization of the viral core with the associated RNA occurs.

- 5 Partial uncoating of the viral core occurs to expose the viral RNA.

Once in the cell cytoplasm, the conversion of the viral RNA into double-stranded DNA commences. In the following replication stage as the viral reverse transcriptase becomes active. Reverse transcriptase synthesizes a double-stranded DNA copy of the single-stranded viral RNA generating a provirus.

- 10 Subsequently, in the integration stage, the viral DNA migrates to and enters the host cell nucleus and becomes integrated into the cell DNA with the help of the enzyme integrase. The provirus can then remain latent or be active, generating products for the generation of new virons. Inside the nucleus, RNA polymerase II transcribes viral DNA into mRNA. The viral mRNA leaves the nucleus.

- 15 A further stage involves a protease activity stage. When viral RNA is translated into a polypeptide sequence, that sequence is assembled in a long chain that includes several individual proteins (reverse transcriptase, protease, integrase). Before these enzymes become functional, they must be cut from the longer polypeptide chain. Viral protease cuts the long chain into its individual enzyme components, which then  
20 facilitates the production of new viruses.

Several of the stages in the above-explained viral replication cycle, may be taken advantage of in developing anti-retroviral therapies. It is known that some antiviral compounds act as inhibitors of HIV on particular stages in the HIV replication cycle  
25 and are effective agents in the treatment of HIV and similar diseases. Fusion inhibitors, for instance, work outside the cell to prevent the first stage of HIV replication. They prevent HIV from entering the CD4 cell by blocking fusion of the outer membrane of the virus with the cell membrane. Another major class of drugs found useful in slowing HIV infections are collectively called "reverse transcriptase inhibitors".

- 30 These act by blocking the recoding of viral RNA into DNA. The reverse transcription stage of HIV replication provides a good target for the action of two types of drugs: nucleoside reverse transcriptase inhibitors (nucleoside analogs) and non-nucleoside

reverse transcriptase inhibitors (NRTIs). Also a number of integrase inhibitors have  
been developed. These include Raltegravir, Elvitegravir, and Dolutegravir. These  
drugs are used in combination with nucleoside reverse transcriptase inhibitors to  
treat HIV infection. The combination of these drugs is highly effective in  
suppressing viral load and preventing disease progression.

Inhibitors work at the last stage of the viral replication cycle. They prevent HIV from being successfully assembled and released from the infected CD4 cell. By blocking the ability of the protease to cleave the viral polypeptide into functional enzymes, protease inhibitors interfere with continued infection.

5

However, despite the fact that various HIV-inhibitors have been identified that are very useful in the treatment of AIDS, they have a common limitation, namely, the targeted enzymes in the HIV virus are able to mutate in such a way that the known drugs become less effective, or even ineffective against these mutant HIV viruses. Or, in other words, the HIV virus creates an ever-increasing resistance against the available drugs. Also, due to an insufficient drug potency in certain cases, incomplete viral suppression may provide fertile ground for resistance to emerge.

10

Thus, due to the great propensity of HIV-1 to mutate, and a continuing growth in the worldwide infection rate, the search for additional therapies for AIDS has been of the highest priority. The emergence of human immunodeficiency virus type 1 (HIV-1) strains resistant to highly active antiretroviral therapy necessitates continued drug discovery for the treatment of HIV-1 infection. It is clear that there is a great need in the art for new antiviral agents to reduce the rate of resistance and suppress viral replication even further.

15

20

In the search for HIV inhibiting compounds, time of addition (TOA) assays are valuable tools for identifying the mechanism of action of test compounds in relation to HIV viral entry, replication, integration and protease activity. Such assays generally comprise the infection of cells with HIV, and the addition of test compounds at various times following infection. Such assays are based on the principle that compound addition at or before the stage in the replication cycle inhibited by the compound results in reduction of virus growth, while compound addition after the stage in the replication cycle inhibited by the compound has no effect.

25

30

However, currently applied time of addition assays have the major disadvantage of being time-consuming and cost-effective. Also, such assays do not sufficiently and accurately permit the distinction between compounds that interfere with particular cellular events within a certain replication stage e.g. an entry, reverse transcription, integration stage, budding and maturation.

35

In view of the above, there is clearly a great need in the art for assays which overcome at least some of the above-mentioned disadvantages. It is therefore an object of the present invention to provide an improved assay for identifying novel drugs acting on particular stages in the HIV replication cycle. It is also an object of the present invention to provide novel anti-retroviral drugs. Furthermore, the invention aims to provide an apparatus for carrying out an improved assay according to the invention.

### Summary

One embodiment of the present invention is a multi-well assay for identifying a compound inhibiting the replication cycle of a micro-organism comprising the subsequent steps of:

- a) preparing a multi-well comprising micro-organism-coated host cells,
- b) initiating at time  $t$  micro-organism infection and replication in said micro-organism-coated host cells such that micro-organism infection and replication is initiated synchronically in all host cells,
- c) bringing at time  $t + \Delta t$  a candidate compound at one or more concentrations into contact with a part of the host cells,
- d) repeating step c) after a time interval of  $\Delta t$  for another part of said host cells,
- e) optionally repeating steps c) and d) using one or more other candidate compounds at one or more concentrations, and
- f) determining whether said candidate compound has inhibited micro-organism replication in said host cells.

Another embodiment of the present invention is an assay as described above wherein said micro-organism is HIV.

Another embodiment of the present invention is an assay as described above, whereby  $\Delta t$  is shorter than the time required for passing from one stage to another stage in the micro-organism replication cycle.

Another embodiment of the present invention is an assay as described above, wherein  $\Delta t$  is shorter than the time required for passing from the entry stage to the reverse transcription stage in the HIV replication cycle.

Another embodiment of the present invention is an assay as described above, wherein  $\Delta t$ , at which compounds are repeatedly added to the multi-well, comprises 1, 2, 3, 4, 5, 10, 15, 20, 25, 30, 40, 60, 120, 240 or 360 minutes.

5 Another embodiment of the present invention is an assay as described above, whereby steps c) to e) are performed under constant reaction conditions (including under a CO concentration of 5 %, a relative humidity comprised between 95 and 100% and a temperature of 37°C).

10 Another embodiment of the present invention is an assay as described above, whereby micro-organism replication in said host cells is initiated at time t by simultaneously bringing all cells at a temperature suitable for initiating micro-organism replication.

15 Another embodiment of the present invention is an assay as described above, whereby said multi-well is prepared by the steps of  
a) infecting host cells with a micro-organism at a high multiplicity of infection,  
b) removing unadsorbed micro-organism, and  
c) bringing said micro-organism-infected host cells onto said well.

20 Another embodiment of the present invention is an assay as described above, whereby said host cells in said multi-well are able to express a gene encoding a detectable protein.

25 Another embodiment of the present invention is an assay as described above, wherein a vector that expresses a gene encoding a detectable protein under the control of a HIV responsive promoter is introduced in said host cells.

30 Another embodiment of the present invention is an assay as described above, wherein said micro-organism is labeled with a detectable protein.

Another embodiment of the present invention is an assay as described above, whereby determination whether said candidate compound is performed by detecting the presence or absence of said detectable reporter.

35

Another embodiment of the present invention is an assay as described above, wherein the presence or the absence of said detectable protein is detected by means of digital imaging techniques.

5 Another embodiment of the present invention is an apparatus for carrying out an assay as described above, comprising:

- a support suitable supporting a multi-well comprising micro-organism - coated host cells, optionally moving in 1 or more directions
- one or more vials for containing a suspension of micro-organism,
- 10 - one or more vials for containing one or more compounds,
- pipetting means for dispensing micro-organisms in said multi-well, optionally moving in 1 or more directions
- pipetting means for dispensing one or more compounds in said multi-well, optionally moving in 1 or more directions
- 15 - pipetting controlling means for controlling dispensing by said pipetting means, and
- environment controlling means for keeping conditions in said apparatus constant while bringing one or more compounds into contact with the host cells.

20

Another embodiment of the present invention is an apparatus as described above wherein said micro-organism is HIV.

25 Another embodiment of the present invention is an apparatus as described above further comprising a temperature-controlled multi-well support.

Another embodiment of the present invention is an apparatus as described above further comprising an insulating cover.

30 Another embodiment of the present invention is an apparatus as described above wherein the circuitry of pipetting and environment controlling means are sealed against high humidity.



Another embodiment of the present invention is a pharmaceutical composition comprising a therapeutically effective amount of one or more compounds identifiable with an assay as described above and a pharmaceutically acceptable excipient.

- 5 Another embodiment of the present invention is a use of a compound, identifiable with an assay as described above, as a medicament.

Another embodiment of the present invention is a use of a compound, identifiable with an assay as described above, for the manufacture of a medicament for treating  
10 infectious diseases.

Another embodiment of the present invention is a method of treating AIDS, comprising administering a therapeutically effective amount of a compound identifiable with an assay as described above to a patient in need thereof.

15 According to the present invention a safe, cost-effective multi-well assay was developed, which enables the analysis of hundreds of thousands of compounds for anti-retroviral activity in an automated and rapid way. In particular, the present assay allow the screening of compounds for antiretroviral activity during the entire HIV-1 life  
20 cycle.

Therefore, in a first embodiment the invention provides a multi-well assay for identifying a compound inhibiting the replication cycle of a micro-organism comprising the subsequent steps of:

- 25 a) preparing a multi-well comprising micro-organism-coated host cells,  
b) initiating at time  $t$ , micro-organism infection and replication in said host cells such that micro-organism infection and replication is initiated synchronically in all host cells,  
c) bringing at time  $t + \Delta t$  a candidate compound at one or more concentrations  
30 into contact with a part of the host cells,  
d) repeating step c) after a time interval of  $\Delta t$  for another part of said host cells,  
e) optionally repeating steps c) and d) using one or more other candidate compounds at one or more concentrations, and  
f) determining whether said candidate compound has inhibited micro-organism  
35 replication in said host cells.

A micro-organism according to the invention may be any of interest that has a replication cycle. Examples include HCV, herpes simples, and preferably AIDS.

5 A first main step in the assay according to the invention comprises the preparation of a multi-well comprising micro-organism infected host cells. The main particularly of this step is that the well is prepared in such a way that micro-organism replication is simultaneously initiated in all micro-organism -infected host cells on the multi-well. This can in particular be obtained by removing unadsorbed micro-organism after interaction of micro-organism with the host cells and prior to transferring the micro-organism -  
10 infected host cells to a multi-well plate. In addition, synchronization of micro-organism replication initiation can be obtained by bringing at a same time  $t$  micro-organism-infected host cells on the multi-well at a temperature suitable for initiating micro-organism replication.

15 Secondly, test compounds are brought into contact with the micro-organism-infected host cells onto the multi-well at different times. The main particularly of this second step is that compounds are added to the multi-well after a time interval  $\Delta t$ , which is shorter than the time required for passing from one stage to another stage in the micro-organism replication cycle.

20 The present assay allows screening of compounds having an antiretroviral activity at various stages of micro-organism replication, e.g. at the entry stage, the reverse transcription stage, the integrase stage and the protease activity stage. Thus, the present assay advantageously permits screening of compounds for antiretroviral activity during the entire micro-organism replication cycle.

25 In addition, as the time interval for adding test compounds to the multi-well are shorter than the time for completing a stage in the replication cycle, the assay permits to screen compounds that interfere with particular events *within* a certain replication micro-organism replication stage.

30 Furthermore, because micro-organism replication is initiated at the same time in all micro-organism-infected host cells on the well, inter-well variation in micro-organism replication is substantially reduced and the assay permits screening of compounds for activity by adding test compounds relatively rapidly into the context of

In a particularly preferred embodiment,  $\Delta t$  is shorter than the time required for passing from the entry stage to the reverse transcription stage in the micro-organism replication cycle. By choosing time intervals  $\Delta t$ , which are shorter than the entry stage of micro-organism in a host cell, the assay enables the identification of compounds that act on particular cellular events that take place during the entry stage of micro-organism replication.

A third main step in the assay comprises rapid and sensitive detection of inhibition of micro-organism replication by tested compounds. Therefore, the micro-organism - infected cells in the multi-well are able to express a gene encoding a reporter protein. Alternatively, also micro-organism may be labeled with a detectable protein. Inhibition by a tested compound is determined by detecting the presence or absence of said reporter protein.

Using the assay according to the present invention thus permits the identification of novel drugs acting on particular stages in the micro-organism replication cycle and also to identify into more detail the specific mode of action of anti-micro-organism drugs within a certain stage in the replication cycle, for instance within the entry stage.

In another embodiment, the present invention relates to an apparatus for carrying out an assay according to the present invention.

The invention further relates to compounds identifiable with an assay according to the invention and pharmaceutical composition comprising a therapeutically effective amount of such compounds. The identified compounds may be used as medicaments, and in particular for the manufacture of a medicament for treating micro-organism infections and in methods for treating micro-organism infections.

Other objects and advantages of the present invention will become apparent from the following detailed description taken in conjunction with the accompanying figures and examples.

#### **Figures**

Figure 1 represents a perspective view of an apparatus according to the present invention.

Figure 2 represents a topical view of an apparatus according to the present invention.

Figure 3 represents the results of an assay according to the invention wherein the effect of reference compounds having known activity on HIV-infected cells are tested at different concentrations.

5 Figure 4 represents the results of an assay according to the invention wherein cell toxicity effects of reference compounds having known activity on HIV-infected cells are tested at different concentrations.

Figure 5 represents the results of an assay according to the invention wherein the effect of reference compounds having known activity on HIV-infected cells are tested at different time points after addition to the HIV-infected cells in a 96-well plate.

10 Figure 6 represents the results of an assay according to the invention wherein the effect of reference compounds having known activity on HIV-infected cells are tested at different time points after addition to the HIV-infected cells in a 384-well plate.

Figure 7a and 7b represent the results of an assay of the invention, wherein the effects of the inhibitor BMS806 upon the activity of HIV-infected cells is evident when the incubation temperatures are 23 deg C (7a) and 37 deg C (7b). The inhibitor is added at regular intervals, starting at time delta t (30 mins, 1, 2, 4, 6, and 8) as indicated in the legend.

15 Figure 8a and 8b represent the results of an assay of the invention, wherein the effects of the inhibitor AMD3100 upon the activity of HIV-infected cells is evident when the incubation temperatures are 23 deg C (7a) and 37 deg C (7b). The inhibitor is added at regular intervals, starting at time delta t (30 mins, 1, 2, 4, 6, and 8) as indicated in the legend.

20 Figure 9a and 9b represent the results of an assay of the invention, wherein the effects of the inhibitor T-20 upon the activity of HIV-infected cells is evident when the incubation temperatures are 23 deg C (7a) and 37 deg C (7b). The inhibitor is added at regular intervals, starting at time delta t (30 mins, 1, 2, 4, 6, and 8) as indicated in the legend.

#### Detailed description of the invention

30 Current HIV therapies involve the use of inhibitors of reverse transcriptase (RT), viral entry and protease enzymes. Despite the development of novel classes of inhibitors and complex drug regimens, drug resistance is increasing. Thus, new types of anti-

Assay

The present invention relates in an embodiment to a multi-well assay for identifying novel compounds which inhibit the HIV replication cycle. The assay provides a convenient model for high-throughput screenings of compounds that show anti-retroviral activity and that can interfere with HIV replication in living cells. Briefly, the present assay comprises three main steps. First, a multi-well comprising HIV-infected host cells is prepared. Secondly, test compounds are brought into contact with HIV-infected host cells applied on the multi-well at different times. A large number of different compounds, optionally at different concentrations, can be applied on the multi-well for testing antiretroviral activities of these compounds. A third step in the assay comprises rapid and sensitive detection of inhibition of HIV replication by the tested compounds.

In particular, the assay comprises the subsequent steps of:

- a) preparing a multi-well comprising HIV-coated host cells under conditions suppressing HIV replication,
- b) initiating at time  $t$  HIV infection and replication in said HIV-coated host cells such that HIV replication is initiated synchronically in all host cells,
- c) bringing at time  $t + \Delta t$  a candidate compound at one or more concentrations into contact with a part of the HIV-infected host cells,
- d) repeating step c) after a time interval of  $\Delta t$  for another part of said HIV-infected host cells,
- e) optionally repeating steps c) and d) using one or more other candidate compounds at one or more concentrations, and
- f) determining whether said candidate compound has inhibited HIV replication in said HIV-infected host cells.

As used herein "human immunodeficiency virus (HIV)" refers to any HIV strain including laboratory HIV strains, wild type HIV strains, mutant HIV strains and any biological sample comprising at least one HIV virus, such as, for example, a HIV clinical isolate. HIV strains compatible with the present invention are any such strains that are capable of infecting mammals, particularly humans. Examples are HIV-1 and HIV-2.

"HIV replication cycle" or "HIV live cycle" or similar terms as used herein refers to the complete cycle passed through by HIV during its replication, starting with contact

and attachment of HIV to a host cell and ending with the production of massive numbers of new viral particles. The HIV replication cycle is meant to encompass various stages, which can be broadly identified as a) an entry stage, b) a reverse transcription stage, c) an integration stage, d) a budding stage and e) maturation (a protease activity) stage.

5  
10  
15  
20  
25  
30  
35  
40  
45  
50  
55  
60  
65  
70  
75  
80  
85  
90  
95  
100  
105  
110  
115  
120  
125  
130  
135  
140  
145  
150  
155  
160  
165  
170  
175  
180  
185  
190  
195  
200  
205  
210  
215  
220  
225  
230  
235  
240  
245  
250  
255  
260  
265  
270  
275  
280  
285  
290  
295  
300  
305  
310  
315  
320  
325  
330  
335  
340  
345  
350  
355  
360  
365  
370  
375  
380  
385  
390  
395  
400  
405  
410  
415  
420  
425  
430  
435  
440  
445  
450  
455  
460  
465  
470  
475  
480  
485  
490  
495  
500  
505  
510  
515  
520  
525  
530  
535  
540  
545  
550  
555  
560  
565  
570  
575  
580  
585  
590  
595  
600  
605  
610  
615  
620  
625  
630  
635  
640  
645  
650  
655  
660  
665  
670  
675  
680  
685  
690  
695  
700  
705  
710  
715  
720  
725  
730  
735  
740  
745  
750  
755  
760  
765  
770  
775  
780  
785  
790  
795  
800  
805  
810  
815  
820  
825  
830  
835  
840  
845  
850  
855  
860  
865  
870  
875  
880  
885  
890  
895  
900  
905  
910  
915  
920  
925  
930  
935  
940  
945  
950  
955  
960  
965  
970  
975  
980  
985  
990  
995

"HIV-infected host cells" as used herein refer to host cells that have been brought into contact with HIV, but wherein replication of HIV has not yet started. Host cells are meant to encompass eukaryotic cells, and preferably mammalian cells from human or animals. In principle, all cells which are susceptible to HIV and which HIV can infect may be used in accordance with the present invention. Preferred host cells for use in assays of the present invention comprise but are not limited to animal or human cells such as CHO, COS, HeLa, C127, 3T3, BHK, HEK 293, Bowes melanoma cells, MT4, D17, etc ...

Although the embodiments described in this section relate to HIV, the method and device of the invention may be applied by the skilled person towards any micro-organism of interest that has different life cycle stages. Examples include HCV, HBV, herpes simplex virus.

The assay according to the invention is performed in multi-well format, which is suitable for screening large amounts of compounds for anti-retroviral activity in an automated and rapid way. The multi-well plates used in the present invention are intended to encompass all sizes of multi-well plates, and preferably comprise 96-well or 384-well plates.

The multi-well plates according to the present invention are prepared by infecting host cells with HIV at a high multiplicity of infection, removing unadsorbed HIV, and bringing said HIV-infected cells onto said well.

Host cells are infected with HIV by bringing the cells into contact with HIV. Preferably the multiplicity of infection (MOI) is higher than 1.0, and more preferably higher. Removing unadsorbed HIV after interaction of HIV with the host cells permits to remove residual HIV that has not interacted with the host cell and which could interfere with the assay.

The infected host cells are then transferred to a multi-well plate. Preferably the concentration of HIV-infected host cells per well is preferably between 10,000 and 600,000 cells per well, and even more preferred comprises 500,000 cells per well.

- 5 In a preferred embodiment, once the HIV-infected host cells have been transferred to a multi-well plate, HIV replication is preferably initiated in all HIV-infected host cells at the same time  $t$  by bringing all host cells on the multi-well at a temperature suitable for initiating HIV replication, being preferably 37°C.
- 10 In a next step, test compounds are added to the multi-well plate comprising the HIV infected cells at various times following initiation of HIV replication. In principle, compound addition at or before a stage in HIV replication, which is inhibited by the compound, results in reduction of virus growth, while compound addition after a step in HIV replication, which is inhibited by the compound, has no effect.

- 15 As used herein "compounds" may refer to known drugs having antiretroviral activity as well as to compounds having unknown antiretroviral activity. The assay according to the invention permits to identify novel compounds having antiretroviral activity as well as to identify into more detail the specific mode of action of readily known anti-HIV
- 20 drugs *during/within* a certain stage in the replication cycle.

- It will be clear from the present invention that several compounds may be tested similarly at one or different concentrations. Time  $t$  according to the invention is the time at which HIV replication is initiated in all cells in the multi-well. After a time  $t + \Delta t$ , a
- 25 compound(s) is transferred to a part of the wells in the multi-well plate. After various additional intervals  $\Delta T$ , the same compound(s) may be transferred to other, not yet treated, wells of the multi-well plate, in order to screen the compound(s) for potential HIV replication inhibiting effects in function of their addition time to the infected HIV cells. This allows evaluating at which particular stage in the HIV cycle the
- 30 compound(s) interfere with HIV replication.

- In addition, since the time intervals  $\Delta t$  is selected in order to be shorter than the time required for passing from one stage to another stage in the HIV replication cycle, the assay allows evaluating at which particular stage in the HIV cycle the compounds
- 35 interfere with HIV replication.

A candidate compound is preferably brought into contact with a part of the HIV-infected host cells in said multi-well at a time  $t + \Delta t$ , whereby  $\Delta t$  is shorter than the time required for passing from one stage to another stage in the HIV replication cycle. According to this embodiment, the present invention permits to screen compounds for antiretroviral activity *within* various stages of the HIV replication cycle, e.g. within the entry stage, the reverse transcription stage, the integrase stage or the protease activity stage. Furthermore, because HIV replication is initiated at the same time in all HIV-infected host cells on the well, the assay permits to test the effect of compounds on HIV replication by adding compounds at relatively rapidly after the onset of HIV replication. The assay thus allows identifying compounds having an antiretroviral activity at very early stages of HIV replication, e.g. compounds that interfere with the entry stage of HIV replication, which is generally performed within 30 minutes post replication initiation. Thus, the present assay advantageously permits screening of compounds for antiretroviral activity at early as well as at late stages during the entire HIV life cycle.

In a particularly preferred embodiment,  $\Delta t$  is shorter than the time required for passing from the entry stage to the reverse transcription stage in the HIV replication cycle. This allows screening for compounds interfering with events taking place during particular phases within the entry stage of HIV replication. Briefly, the entry stage of HIV replication distinguishes a CD4 receptor attachment phase, a co-receptor binding phase and a phase involving membrane fusion events. By choosing time intervals  $\Delta t$ , which are shorter than the entry stage of HIV in a host cell, the assay enables the identification of compounds that have anti-retroviral activity during one or more of these above-referred phases of the entry stage. In particular, the present invention so allows the identification of compounds interfering with either the CD4 receptor attachment phase, the co-receptor binding phase and/or the membrane fusion phase. It is clear that compounds having antiretroviral activity at very early stages in HIV replication, in particular even at particular phases during the entry stage of HIV, are of particular importance for anti-HIV therapies, since rapid inhibition of HIV replication increases the chances of successfully treating HIV.



In another preferred embodiment, reaction conditions applied during the addition of compounds to the multi-well in steps c) to e) of the assay are kept constant.

5 In one embodiment of the invention, the temperature T of the wells is determined by the user. For example, a temperature of 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50 or higher might be selected, and is kept constant throughout out the experiment.

10 In one embodiment of the invention, the temperature of every well in the multiwell is the same.

15 In one embodiment of the invention, the temperature of at least one well is different from another. For example, a temperature gradient might be held across the width of the multi-well plate such that the left side of the plate is hotter than the right side of the plate. Alternatively, the temperature of an individual well in a multiwell plate may be different.

20 In another embodiment of the invention, a means is present to prevent the contents of the wells from reducing in volume by evaporation. For example, a relative humidity of between 95 and 100 % is provided. Alternatively, a removable heated lid is provided which is applied on top of the multiwell plate between additions of reagents.

25 Preferably, the following conditions are maintained: a CO concentration of 5 %, a relative humidity comprised between 95 and 100% and a temperature of 37°C.

30 Inhibition of HIV replication by a compound is determined by detecting the presence or absence of said detectable protein, which is released either by the host cell or by the virus as such.

35 In one embodiment, HIV-infected cells in the multi-well are engineered in order to be capable of expressing a gene encoding a detectable (reporter) protein. Preferably, a gene encoding a detectable protein is put under the control of a HIV responsive promoter and introduced in said host cells. The reporter construct can be transferred

5 The term "HIV responsive promoters" as used in the present invention is meant to include any transcriptional control unit capable of initiating transcription upon initiation of virus replication and includes regulatory elements such as enhancers and other regions binding transcriptional control factors. Examples of suitable HIV responsive promoters are known in the art.

Suitable genes encoding reporter proteins according to the present invention may be any reporter gene known to the skilled person as being active in cells. The reporter gene preferably encodes an enzyme which catalyses a reaction or may be a protein which produces a detectable signal, preferably a visually detectable signal, such as a colored product. Examples of suitable reporter genes comprise but are not limited to genes encoding chloramphenicol acetyltransferase (CAT),  $\beta$ -galactosidase (GAL),  $\beta$ -glucuronidase (GUS), luciferase (LUC), green fluorescent protein (GFP), etc ..

In another embodiment, HIV is labeled with a detectable protein. Examples methods of labeling an organism using a detectable protein are known in the art and include, for example, the use of lyptotracers, VRP-GFP or any suitable GFP-fusion (Mc Donald *et al*, *JCB* 159, 3, pp441-452).

In one embodiment, the detectable protein is detected *directly*, in case of a colored, fluorescent or luminescent protein. Fluorescence, for example that is produced as a result of luciferase activity, may be quantitated using a spectrophotometer.

The presence and/or amount of gene product resulting from expression from the reporter gene may also be determined *indirectly* by using immunolabelling techniques.

The term "immunolabelling technique" as used herein is meant to refer to various detection methods that use immunoglobulins to detect specific epitopes. Such immunolabelling techniques may comprise but are not limited to ELISA.

<sup>a</sup> The number of subjects who were included in each group was determined by the number of subjects who completed the study.

Detection is preferably performed by means of digital imaging techniques. Such techniques are known in the art and include, for example, the use of a luminometer, scanning microscope, spectrophotometer, and preferably a fluorescence reader.

5

Apparatus

In another embodiment, the invention relates to an apparatus for carrying out an assay according to the present invention. The apparatus 1, of which an example is illustrated in FIG. 1 and FIG. 2, comprises

- 10       - a support suitable for moving in 1 or 2 directions for supporting a multi-well comprising HIV-infected host cells,
- one or more vials for containing one or more compounds,
- pipetting means for dispensing one or more compounds in a multi-well, positionable on said support
- 15       - controlling means for controlling dispensing by said pipetting means
- controlling means for controlling the reaction conditions in said apparatus
- controlling means for temperature
- optionally controlling means for humidity

- 20       As illustrated in FIG.1 and FIG 2 an apparatus is represented comprising a support 2 whereon a multi-well comprising HIV-infected host cells can be positioned. The support is preferably movable, in 1 or 2 directions, e.g. in X1 and Y1 direction. Therefore, the support 2 can for instance be positioned on a X-Y table 6. Such X-Y tables are well known in the art and not described into detail in the present
- 25       specification.

Alternatively, the support 2 may be stationary, and an arm for liquid dispensing 4 is movable in 1 or more directions e.g. in Y1 and optionally in X1.

- 30       Alternatively, both an arm as described above and an X-Y table may be movable according to a device of the invention.

- 35       In another embodiment of the invention, the plate support is heated so as to maintain the temperature of the multiwell plate according to the method. It is an aspect of the invention, that the plate support has the means to maintain the same and constant temperature across its surface. It is another aspect of the invention, that the plate

support has the means to maintain a constant temperature gradient across its surface (e.g. a temperature gradient from left to right). It is another aspect of the invention, that the plate support has the means to maintain a constant and different temperature in each well of the multiwell plate.

5

In another embodiment of the present invention, the control of the temperature is optimized such that the entire area is kept constant in temperature. If necessary, the temperature can be incrementally increased or decreased. Examples of temperatures are 4°C, room temperature ( $\pm 22^\circ\text{C}$ ) or 37°C.

10

In a preferred embodiment, the apparatus comprises one or more containers holding the compounds to be tested. The container may be one or more multi-well plates, vials or container known in the art. The transfer of the compounds to the multi-well plate, which is to be positioned on the support 2, can be performed by pipetting means 4.

15 Preferably, these pipetting means 4 are movable in three directions, X, Y and Z.

The present apparatus further comprises one or more controlling means. A control means 5 is provided for controlling the reaction conditions in the apparatus, before as well as after initiation of HIV replication. At the beginning of the assay, the conditions of the apparatus should be regulated such that HIV replication is suppressed. Only at 20 time t, HIV replication should be initiated for all cells in the multi-well at the same time by controlling the temperature in the apparatus. In addition, the reaction conditions in the incubator should be kept constant after initiation of replication and during an after addition of compounds to be tested, such that all compounds can be screened under 25 similar reaction conditions. Therefore, the controlling means monitors and maintains parameters such as plate temperature, ambient air temperature, ambient air humidity, and the gaseous composition the ambient air.

It is an aspect of the invention that the apparatus is enclosed in an environment in 30 which the ambient air is controlled. For example of level of CO<sub>2</sub>, the temperature and/or humidity may be controlled by the user. It is an aspect of the invention that the electronics are provided in the same environment as the multiwell plate and the apparatus is sealed to be of substantially airtight environment and to have a constant humidity.

It is an aspect of the invention that the environment is enclosed by means cover that seal the device substantially from the environment of the user. Said cover is part of the device. The cover may be partially transparent to enable the user to see the multiwell plate. The cover may be have a insulative properties enable so as to stabilize the temperature. The cover may be have be airtight, so as to stabilize the level of CO<sub>2</sub>.

Further control means are preferably provided for controlling the pipetting action of the pipetting means 4. This control means may be similar or different from the control means 5 for regulating the reaction conditions in the apparatus. The pipetting control means are suitable for determining and regulating the intervals  $\Delta t$  at which compounds need to be added to the multi-well.

It is an aspect of the invention that the control means, and the pipettor controlled thereby are capable of dispensing nanolitre volumes. Examples of automatic dispensing devices may be any of the art. They include those capable of delivering down to nonalitre volumes and may be operated by micro pumps, and piezo pumps.

In another embodiment of the invention, the apparatus is further provided with a heated lid which is brought into contact with the top surface of the multiwell plate during incubations. The temperature of well-side of the lid is maintained sufficiently high to prevent the contents of the well from evaporating. The lid may have a means to seal with the surface of the multiwell plate, such as a gasket. The lid is mechanically be brought into contact with the top surface of the multiwell plate during incubations. During an addition operation, the lid is mechanically opened. The lid may be mechanically opened by any mechanised means, such as, for example, a robot arm, a motorized hinge etc.

The apparatus according to the invention has the advantage of enabling compound testing at various time intervals  $\Delta t$ , without the need of taking the multi-well plate out of the apparatus every time compounds need to be added to the multi-well. This can be obtained by providing all necessary equipment for screening compounds for antiretroviral activity within apparatus 1, and by providing means 5 which permit to control and set up constant reaction conditions in the apparatus.

In another preferred embodiment, the apparatus is further provided with detecting means for determining whether the tested compounds have inhibited HIV replication. These detecting means may for instance comprise imaging means.

- 5 In yet another preferred embodiment, the apparatus is further provided with analyzing means for analyzing the effects of tested compounds on HIV replication. These analyzing means may for instance comprise computer means, suitable for registering and determining at which stage replication, or at which phase within a replication stage the compounds show antiretroviral effects.

10

- In another preferred embodiment, the apparatus according to the invention preferably comprises different units, including a loading unit, wherein test compounds are transferred to the multi-well plates, a detection unit, wherein potential inhibitory effects of tested compounds are determined, and an analysis unit, wherein the effects of the test compounds are analyzed. Preferably, the apparatus comprises transport means, for transporting the multi-well plates from one unit to another in said apparatus.

15

#### Medicinal uses

- 20 It was found that the present assay provides the necessary sensitivity for identifying antiviral molecules. Thus, the assay according to the invention can be utilized to facilitate the development of a safe, efficient method for screening compound libraries for anti-HIV activity.

#### Reach through products

- 25 In a further aspect the invention relates to compounds identifiable with an assay according to the invention. The compounds are particularly suitable for being applied in AIDS therapies. The term "AIDS (acquired immunodeficiency syndrome)" refers to the late stage of HIV disease, characterized by a deterioration of the immune system and a susceptibility to a range of opportunistic infections and cancers.

30

In another further embodiment, a pharmaceutical composition is provided comprising a therapeutically effective amount of one or more compounds identifiable with an assay according to the invention and at least one pharmaceutically acceptable excipient.

according to the invention as a medicament or for the manufacture of a medicament for treating AIDS.

5 In yet another embodiment, the invention relates to a method of treating AIDS, comprising administering a therapeutically effective amount of a compound identifiable with an assay according to the invention to a patient in need thereof.

10 The following examples are meant to illustrate the present invention. These examples are presented to exemplify the invention and are not to be considered as limiting the scope of the invention.

### **Examples**

#### ***Example 1***

15 In a time-of-addition experiment according to the present invention, the step in the HIV replication cycle in which a compound is active is determined and compared with reference compounds including inhibitors for binding/fusion (entry process), reverse transcriptase, integrase and protease. When a potent antiviral compound is added at the time of infection, no viral replication takes place. But, if addition of compound is delayed, protection can be observed up to the moment that the virus passes the stage  
20 at which the inhibitor interacts. The use of reference compounds with a known mode of is essential for the correct interpretation of the results.

A MT4 cell line expressing EGFP reporter gene under the control of a HIV-1-LTR promoter element can be used. MT4-LTR-EGFP cells are infected at a high multiplicity  
25 of infection (MOI) by centrifugation for 10 min at 400 g. Unadsorbed virus is then removed by two washing steps at 4°C in order to synchronize the infection. Multi-well plates loaded with cells are then incubated at 37°C, being a temperature suitable for initiating the HIV replication cycle.

30 From 5, 10, 15, 20, 30 or more min post-infection on, compounds to be tested are added to parallel cultures in micro-titer plates at different times. The cultures can be scored microscopically for fluorescence 24 hours after infection and supernatant can be collected. HIV replication in the supernatant samples can be quantified by measuring the concentration of the p24 viral antigen using a commercial kit, according  
35 to the manufacturer protocol (NEN).

### **Example 2**

The present example illustrates a method according to the invention for identifying compounds having an inhibiting effect on HIV during the entry process of the HIV replication cycle.

A MT4 cell line was used in the present example expressing a luciferase reporter gene under the control of a HIV-1-LTR promoter element. Cells were counted and a cell suspension of 500.000 ml or 350.000 ml was made for use in 96-well or 384 wells, respectively. The cells were centrifuged for 10 minutes at 400g. Subsequently, cells were infected with at high multiplicity of infection with HIV-1 (strain IIIB) at a host cell/virus titer of 1/1. Unabsorbed virus was removed by two washing steps in order to synchronize the infection. After the second washing step, cells were resuspended and the suspensions were distributed in the wells of a micro-titer plate in an amount of 95µl or 36µl in a 96-well or a 384 well format, respectively. As compounds to be tested, reference compounds having well known mechanisms of action were included in the plates.

#### **2.1 Titration of compounds under TOA-conditions**

96-well plates loaded with cells coated with viruses as explained above were incubated at 37°C, being a temperature suitable for initiating the HIV replication cycle. 30 minutes post-infection; a dilution series of compounds to be tested was added to the wells of the micro-titer plate in an amount of 5 µl/well. The plates were further incubated at 37°C for 26 hours.

As compounds to be tested, reference compounds having well known mechanisms of action were included in the plates. The reference compounds comprised:

- ATA, AMD 3100 and T20 having inhibiting effects on the HIV entry process,
- EFV having inhibiting effects on the HIV replication process,
- TMC143205 having inhibiting effects on the HIV integration process,
- SQV having inhibiting effects on the HIV protease activity process.

TA were used in a concentration from 5µM to 1000-3 µM while all other compounds were used in a concentration from 1 µM to 1000-3 µM.



Toxicity measurements, with the aim of determining compounds that demonstrate the most activity for the least toxicity were performed in parallel with activity testing of the cited compounds. Aspecific activity of the compounds was measured by incubating the same cells as described above using the same protocol as described above, without  
5 addition of virus particles.

In order to determine whether said test compounds had inhibited HIV replication in the host cells, the presence or absence of the luciferase gene product was measured. Therefore, 100µl Luc-lit<sup>TM</sup> substrate was added to each well and luminescence was  
10 monitored by processing the plates in a PerkinElmer-Viewlux<sup>TM</sup> within 30 minutes. HIV replication was quantified by measuring the luciferase signal.

Dose response curves for activity and toxicity are presented in Fig. 3 and Fig. 4 respectively. From the figures it can be seen that the highest compound concentration  
15 used in the assays exhibited no toxicity (Figure 4) and gave approximately 100 % inhibition (Figure 3). The results demonstrate that the method of the invention enable the selection of compounds which exhibit the most activity for the least toxicity.

#### 2.2 Time aspect of the method according to the invention using luminescent cells

20 96-well plates prepared as indicated above were incubated at 37°C, being a temperature suitable for initiating the HIV replication cycle. 30 minutes post-infection; a dilution series of compounds to be tested was added to the wells of the micro-titer plate in an amount of 5 µl/well at different time points. The plates were further incubated at 37°C for 26 hours.

25

Also 384-well plates, prepared as indicated above, were incubated at 37°C, being a temperature suitable for initiating the HIV replication cycle. 30 minutes post infection; compounds to be tested were added to the wells of the micro-titer plate in an amount of 4 µl/well. The plates were further incubated at 37°C for 26 hours.

30

The reference compounds as indicated in 1.1 were tested. ATA was used in a concentration of 5µM; AMD3100, T20 EFV ad SQV were used in a concentration of 1µM and TMC143205 was used in a concentration of 1µM. In this example, the time points for adding the compounds comprised 30, 45, 60, 75, 90, 105, 120, 135, 150,  
35 165, 180, 210, 240, 270, 300, 330, 360, 390, 420, 450, 480, 510, 540 and 570 minutes. 100µl Luc-lit<sup>TM</sup> substrate was added to each well and luminescence was

monitored by processing the 96-well plates in a PerkinElmer-Viewlux<sup>TM</sup> and the 384-well plates in a Wallac-Viewlux<sup>TM</sup> within 30 minutes. HIV replication was quantified by measuring the luciferase signal.

5 Dose response curves for activity of tested compounds in 96-well plates and in 384-well plates are presented in Fig. 5 and Fig. 6, respectively. From the figures it can be seen that all reference compounds behaved as expected. Similar results are obtained on 96-well plates and 384-well plates. More in particular, it is clear that the entry inhibitors ATA, AMD3100 and T20 all demonstrated a strong inhibitory activity in the  
10 early stages of addition.

Example 2 illustrates that the present method allows detecting compounds having an effect on the HIV entry process.

15 ***Example 3***

Experiments were performed according to the invention, where the effects of different inhibitors, at different concentrations and delta t's upon HIV infected cells were examined at two different temperatures. Experimental detail is provided in the Figure legends 7 to 9.

20

The results demonstrate the effect of temperature upon the inhibitory activity of various HIV inhibitors, and show the clear advantage of the method of the invention.

## Claims

1. Multi-well assay for identifying a compound inhibiting the replication cycle of a micro-organism comprising the subsequent steps of:
  - a) preparing a multi-well comprising micro-organism-coated host cells,
  - b) initiating at time  $t$  micro-organism infection and replication in said micro-organism-coated host cells such that micro-organism infection and replication is initiated synchronically in all host cells,
  - c) bringing at time  $t + \Delta t$  a candidate compound at one or more concentrations into contact with a part of the host cells,
  - d) repeating step c) after a time interval of  $\Delta t$  for another part of said host cells,
  - e) optionally repeating steps c) and d) using one or more other candidate compounds at one or more concentrations, and
  - f) determining whether said candidate compound has inhibited micro-organism replication in said host cells.
2. Assay according to claim 1 wherein said micro-organism is HIV.
3. Assay according to claims 1 and 2, whereby  $\Delta t$  is shorter than the time required for passing from one stage to another stage in the micro-organism replication cycle.
4. Assay according to any of claim 1 to 3, whereby  $\Delta t$  is shorter than the time required for passing from the entry stage to the reverse transcription stage in the micro-organism replication cycle.
5. Assay according to any of claims 1-4, wherein  $\Delta t$ , at which compounds are repeatedly added to the multi-well, comprises 1, 2, 3, 4, 5, 10, 15, 20, 25, 30, 40, 60, 120, 240 or 360 minutes.
6. Assay according any of claims 1-5, whereby steps c) to e) are performed under constant reaction conditions (including under a  $\text{CO}_2$ -concentration of 5 %, a relative humidity comprised between 95 and 100% and a temperature of  $37^\circ\text{C}$ ).

7. Assay according to any of claims 1 to 6, whereby micro-organism replication in said host cells is initiated at time  $t$  by simultaneously bringing all cells at a temperature suitable for initiating micro-organism infection and replication.
- 5 8. Assay according to any of claims 1 to 7, whereby said multi-well is prepared by the steps of
- a) infecting host cells with a micro-organism at a high multiplicity of infection,
  - b) removing unadsorbed micro-organism, and
  - c) bringing said micro-organism-infected host cells onto said well.
- 10 9. Assay according to any of claims 1-8, whereby said host cells in said multi-well are able to express a gene encoding a detectable protein.
- 15 10. Assay according to claim 9, wherein a vector that expresses a gene encoding a detectable protein under the control of a HIV responsive promoter is introduced in said host cells.
11. Assay according to any of claims 1-10, wherein said micro-organism is labeled with a detectable protein.
- 20 12. Assay according to any of claims 1-11, whereby determination whether said candidate compound is performed by detecting the presence or absence of said detectable reporter.
- 25 13. Assay according to claim 12, wherein the presence or the absence of said detectable protein is detected by means of digital imaging techniques.
14. Apparatus for carrying out an assay according to the any of claims 1-13, comprising:
- 30 - a support suitable supporting a multi-well comprising micro-organism - coated host cells, optionally moving in 1 or more directions
- one or more reservoirs for containing a suspension of micro-organism.
- one or more reservoirs containing one or more compounds
- one or more reservoirs containing one or more compounds
- one or more reservoirs containing one or more compounds
- one or more reservoirs containing one or more compounds

- pipetting means for dispensing one or more compounds in said multi-well, optionally moving in 1 or more directions
  - pipetting controlling means for controlling dispensing by said pipetting means, and
  - 5       - environment controlling means for keeping conditions in said apparatus constant while bringing one or more compounds into contact with the host cells.
- 10       15. An apparatus according to claim 14 wherein said micro-organism is HIV.
16. An apparatus according to claims 14 and 15 further comprising a temperature-controlled multi-well support.
- 15       17. An apparatus according to any of claims 14 to 16 further comprising an insulating cover.
18. An apparatus according to claims 14 to 17 wherein the circuitry of pipetting and environment controlling means are sealed against high humidity.
- 20       19. Compounds identifiable with an assay according to any of claims 1 to 13.
20. Pharmaceutical composition comprising a therapeutically effective amount of one or more compounds identifiable with an assay according to any of claims 1 to 13 and a pharmaceutically acceptable excipient.
- 25       21. Use of a compound, identifiable with an assay according to any of claims 1 to 13, as a medicament.
22. Use of a compound, identifiable with an assay according to any of claims 1 to 13, for the manufacture of a medicament for treating infectious diseases.
- 30       23. Method of treating AIDS, comprising administering a therapeutically effective amount of a compound identifiable with an assay according to any of claims 2 to 13 to a patient in need thereof.
- 35

**Abstract**

**5 Multi-well assay for identifying the inhibition activity of compounds acting at different stages of microbial replication and having anti-microbial activity**

The present invention relates to a multi-well assay for identifying a compound inhibiting the replication cycle of a micro-organism, for example, HIV, and to an apparatus for carrying out an assay according to the present invention. The invention further relates to compounds identifiable with an assay according to the invention and pharmaceutical composition comprising a therapeutically effective amount of such compounds. The identified compounds may be used as medicaments, and in particular for the manufacture of a medicament for treating infectious diseases and in  
**10**  
**15** method for treating infectious diseases including AIDS.

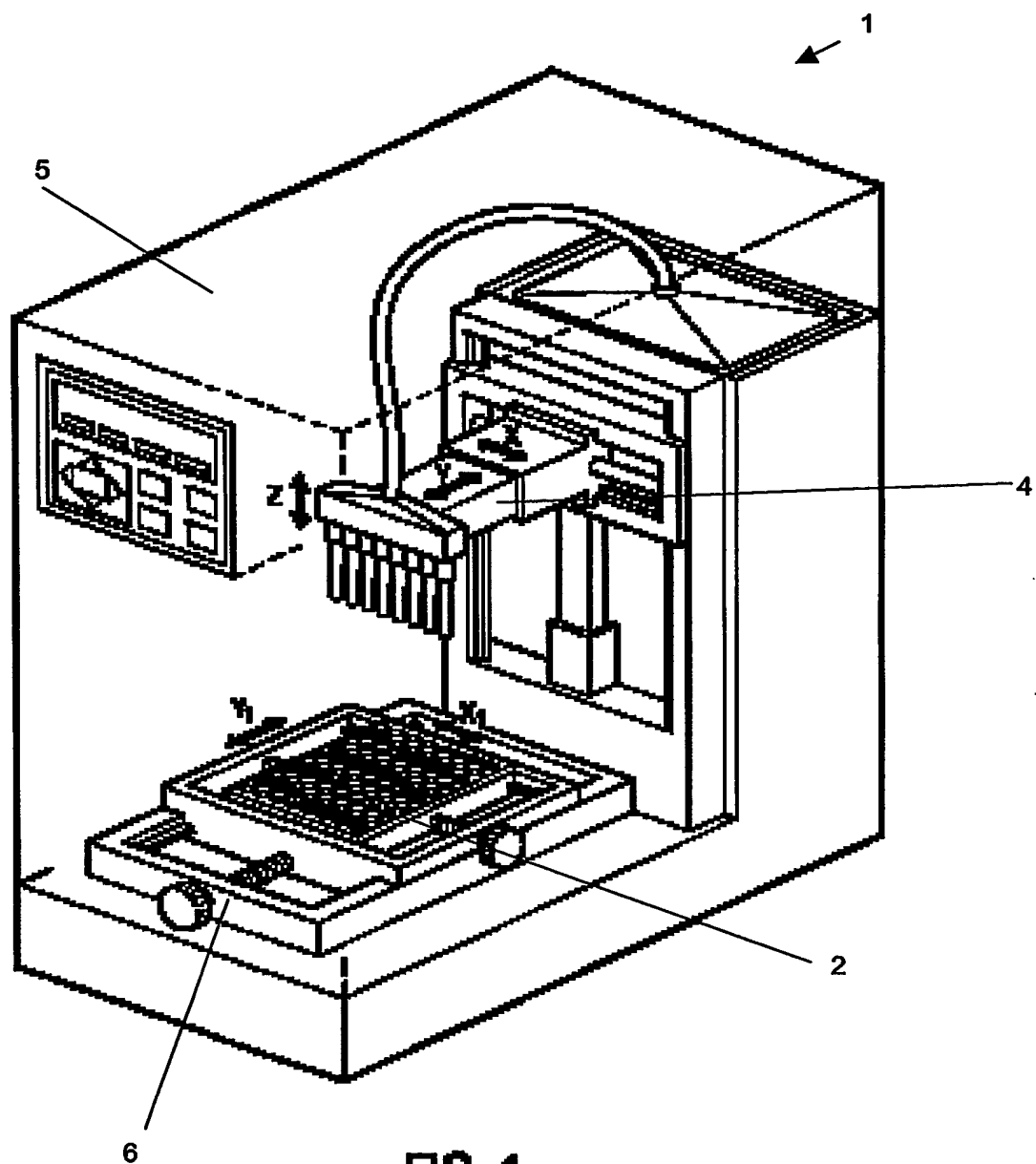


FIG. 1

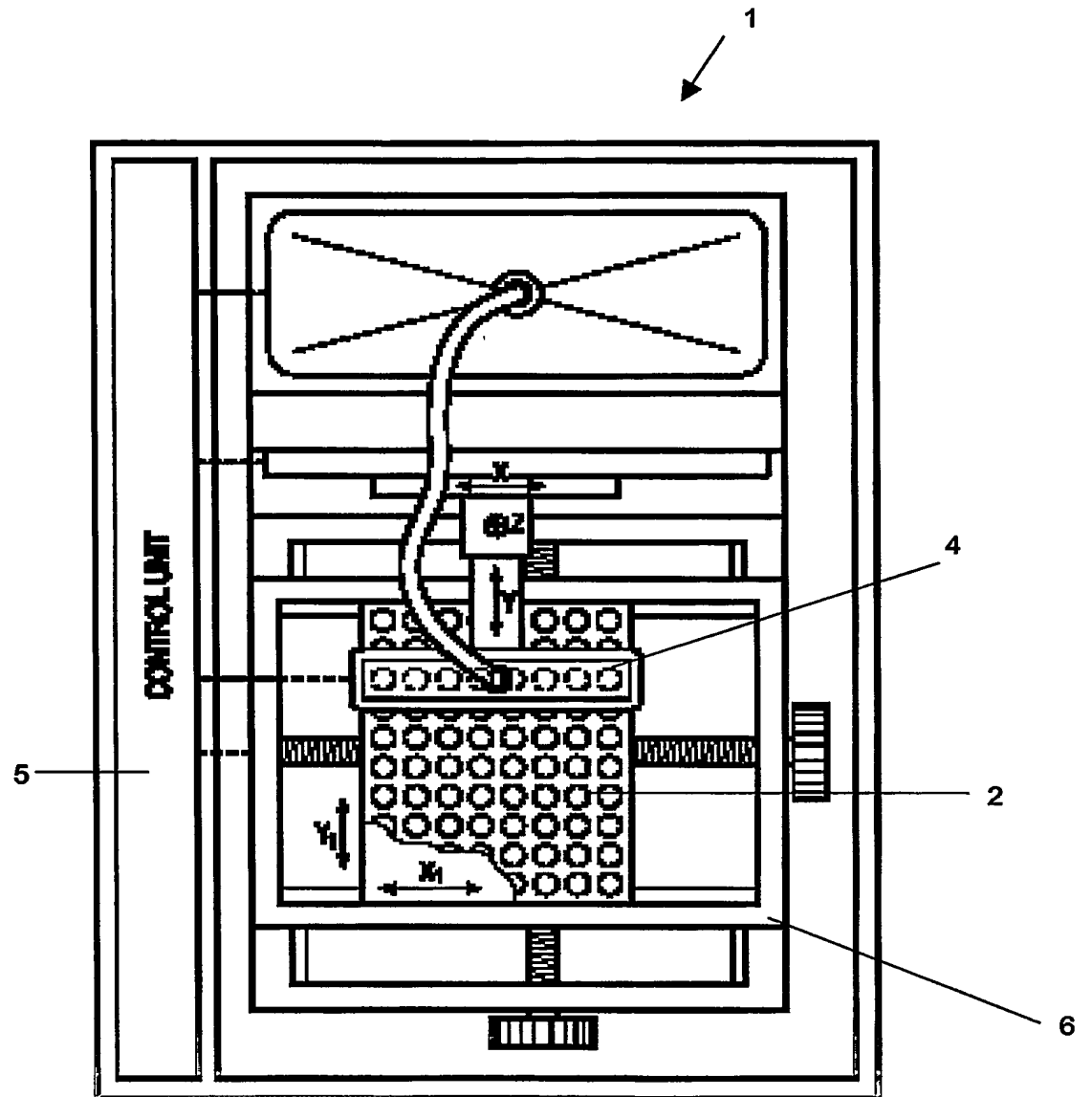


FIG. 2



3/10

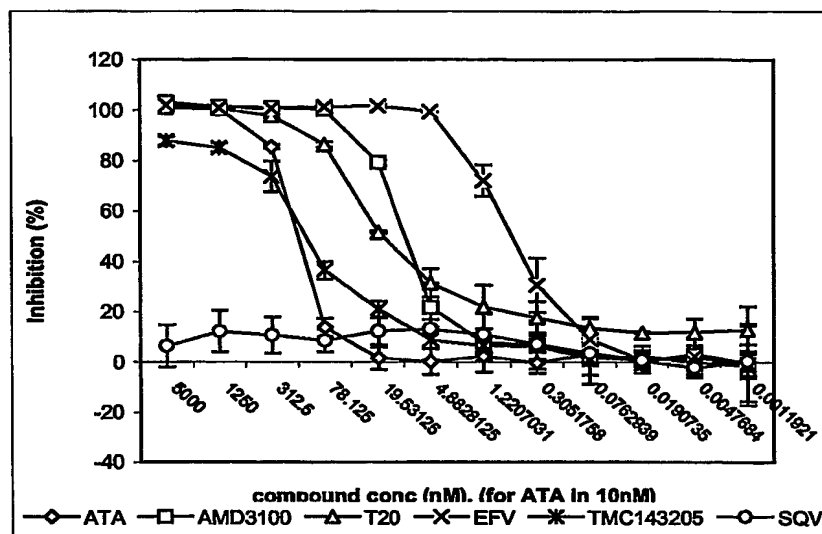


Fig. 3

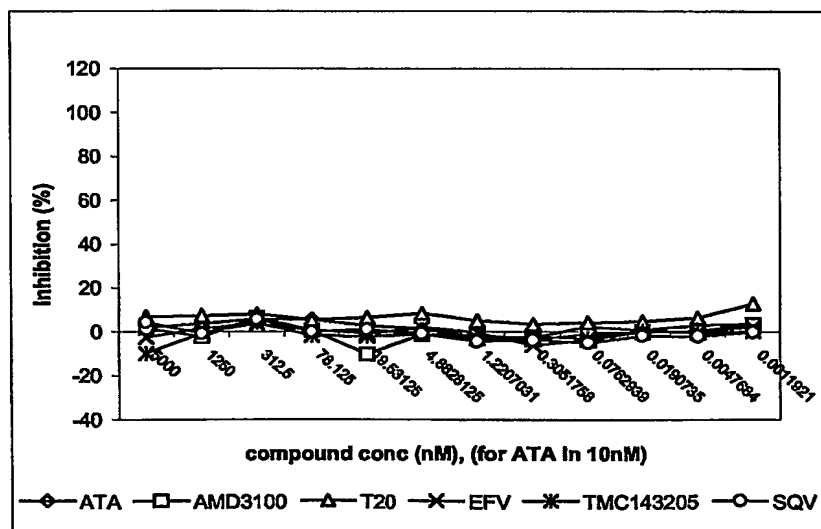


Fig. 4

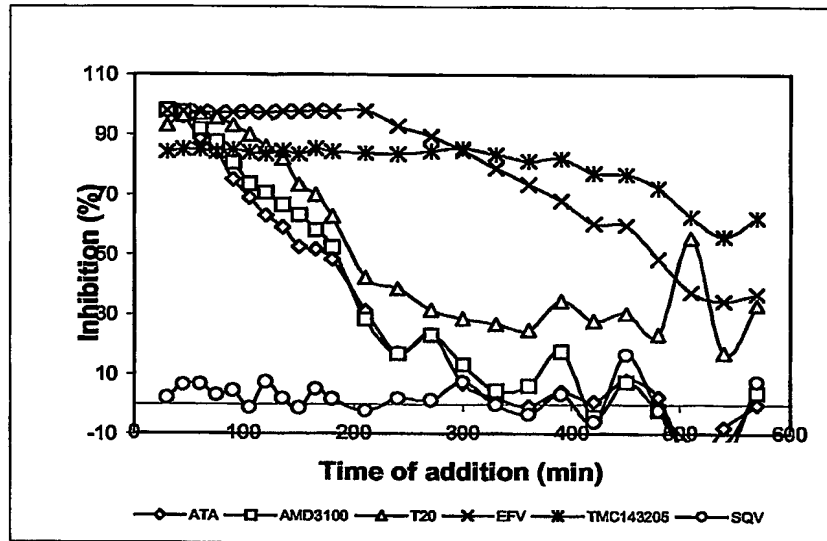


Fig. 5

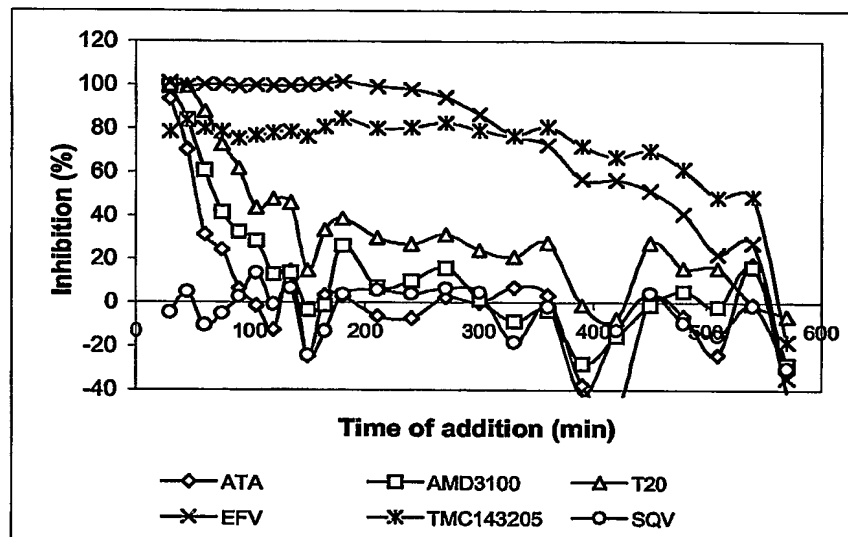
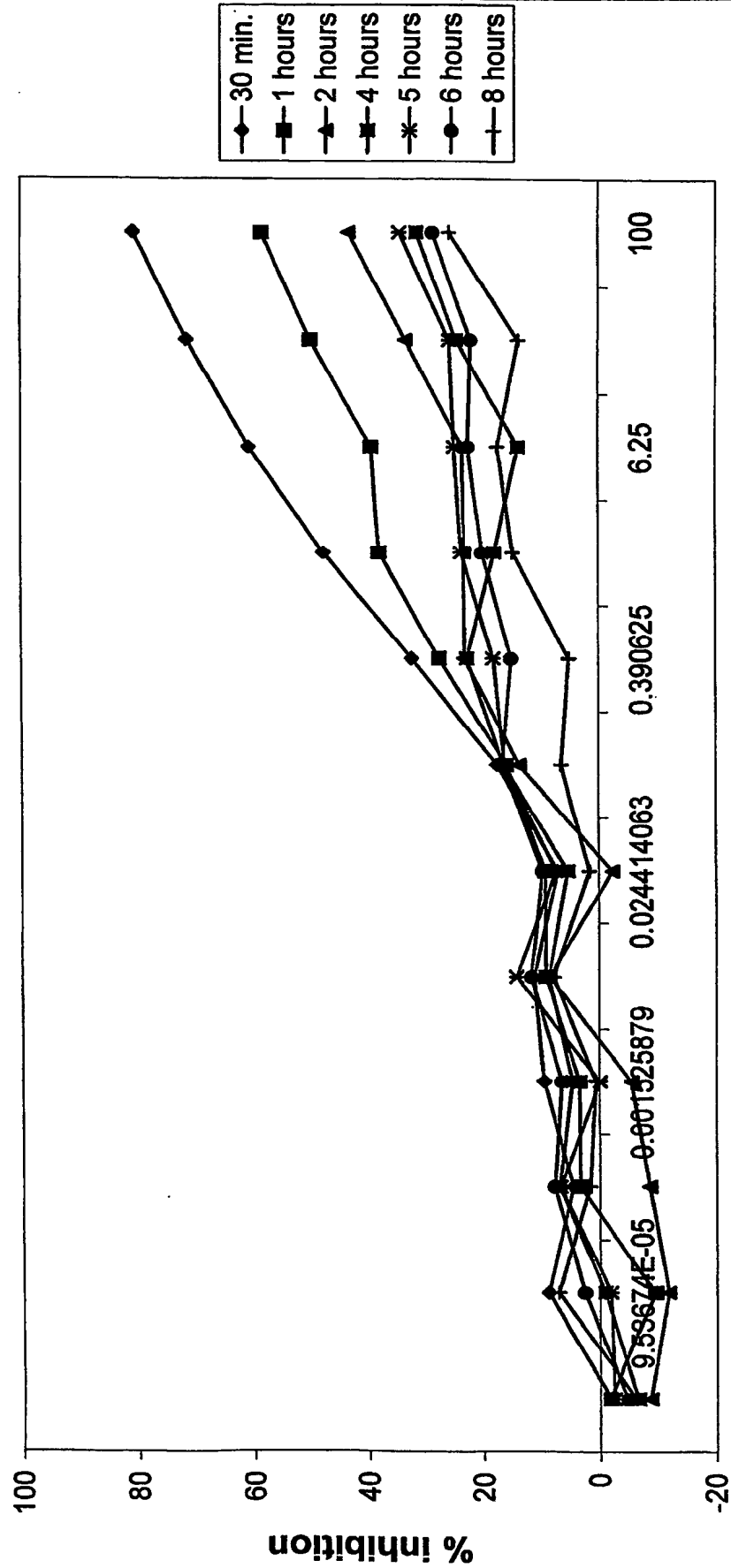


Fig. 6

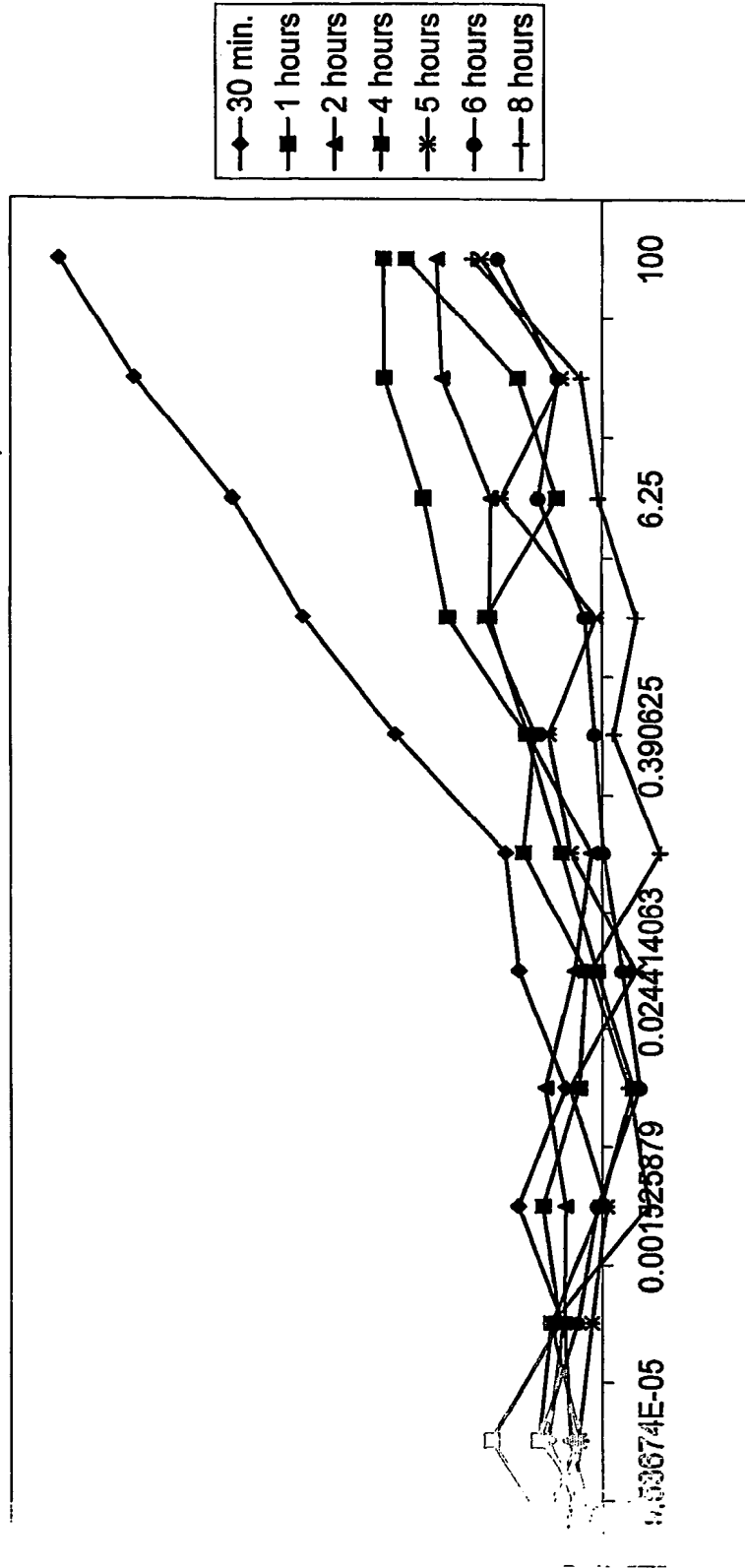
# Anti-viral-experiment with TOA protocol: 23°C



Concentration of BMS806 ( $\mu\text{M}$ )

Figure 7A

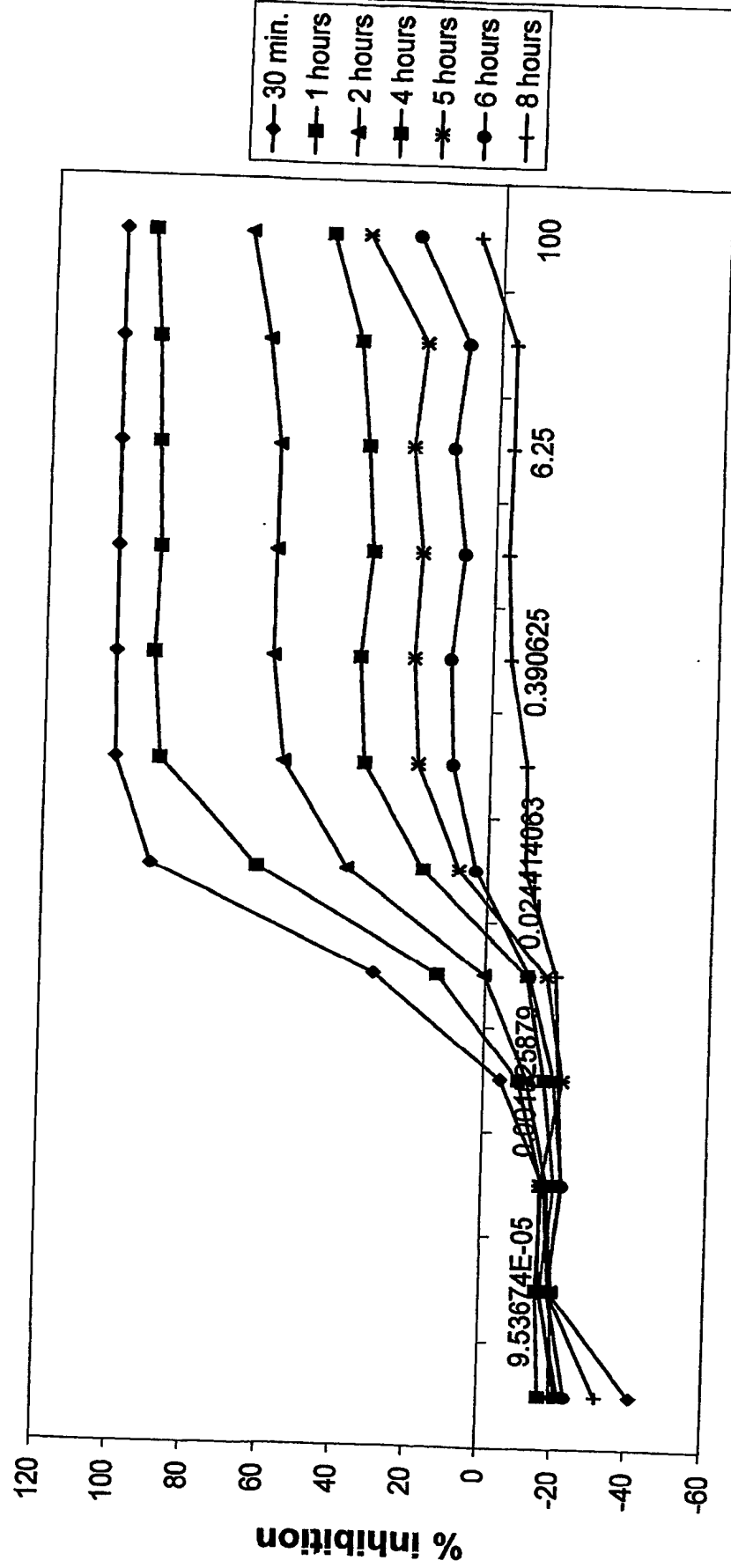
# Anti-viral-experiment with TOA protocol: 37°C



Concentration of BMS806 (µM)

Figure 7B

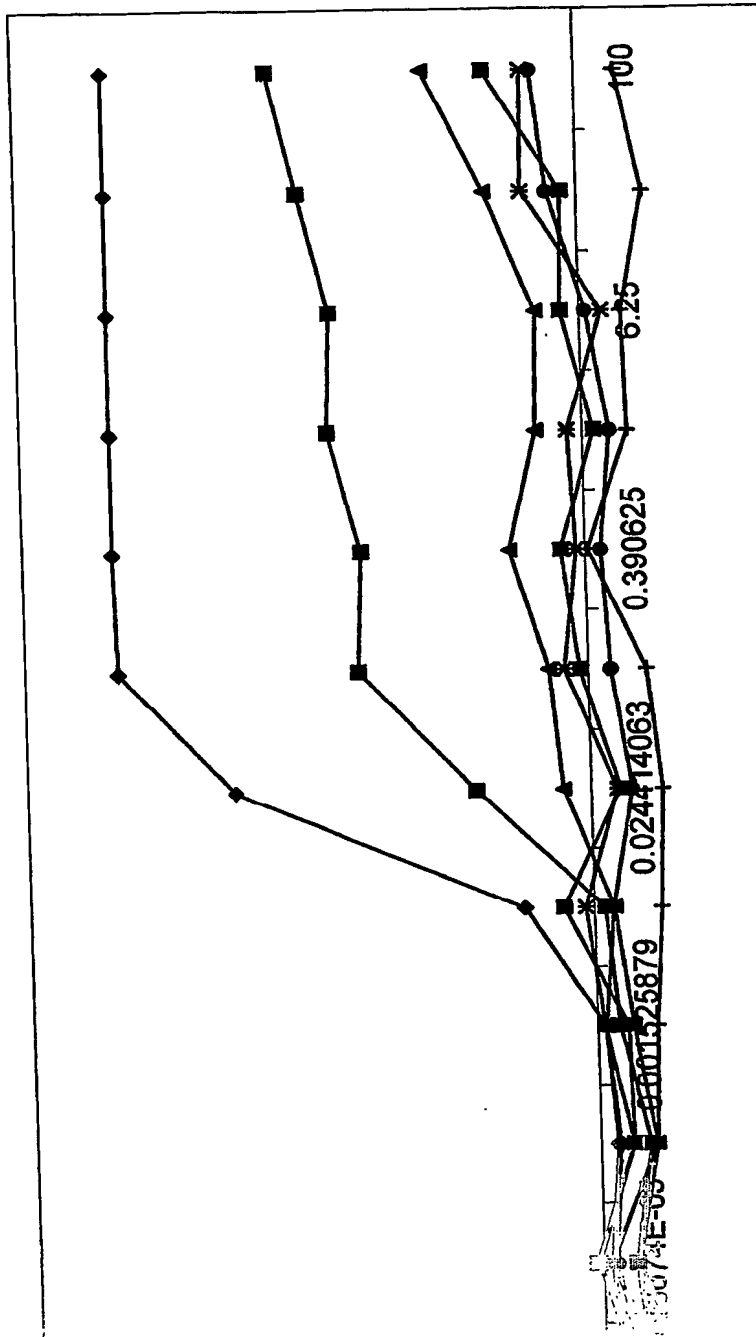
# Anti-viral-experiment with TOA protocol: 23°C



Concentration of AMD3100 (µM)

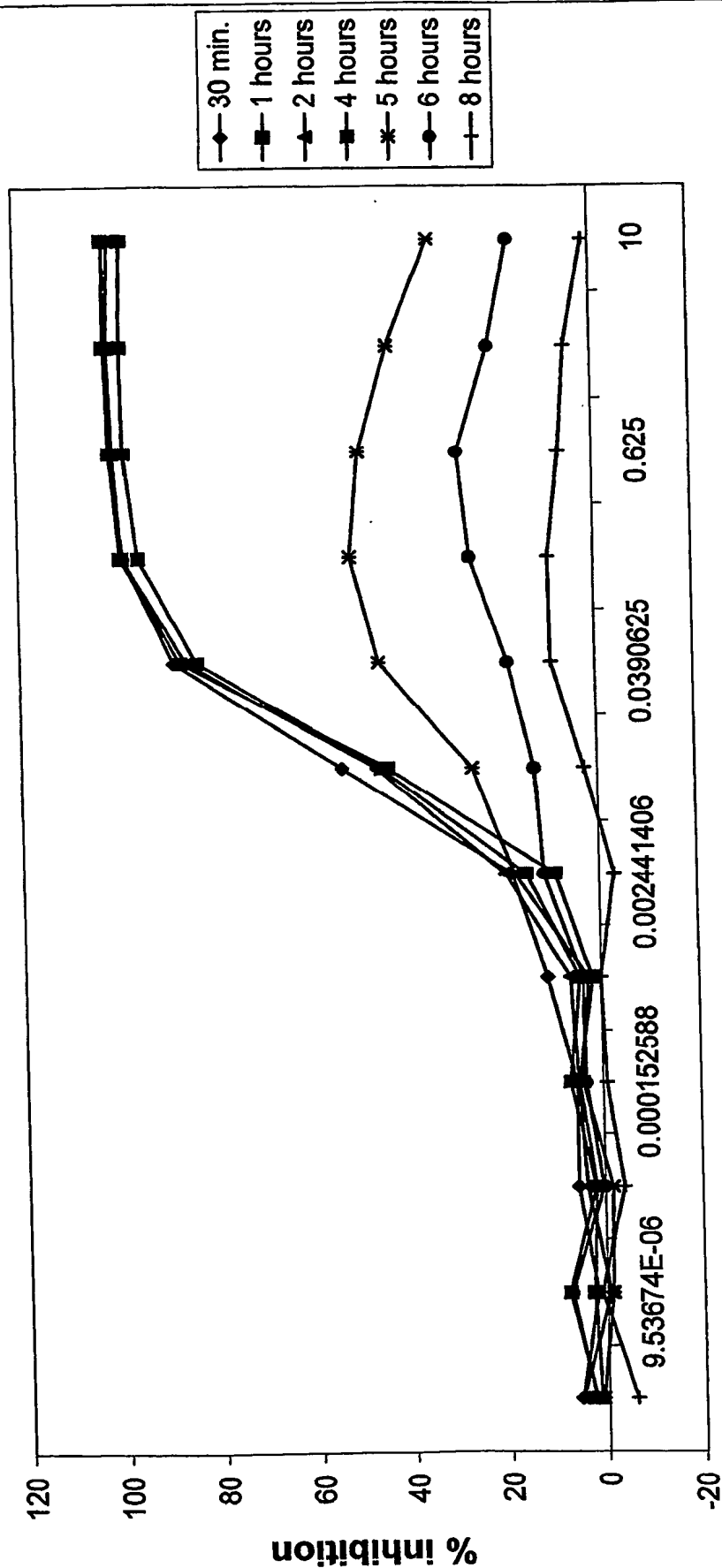
Figure 8A

# Anti-viral-experiment with TOA protocol: 37°C

Concentration of AMD3100 ( $\mu\text{M}$ )

## Figure 8B

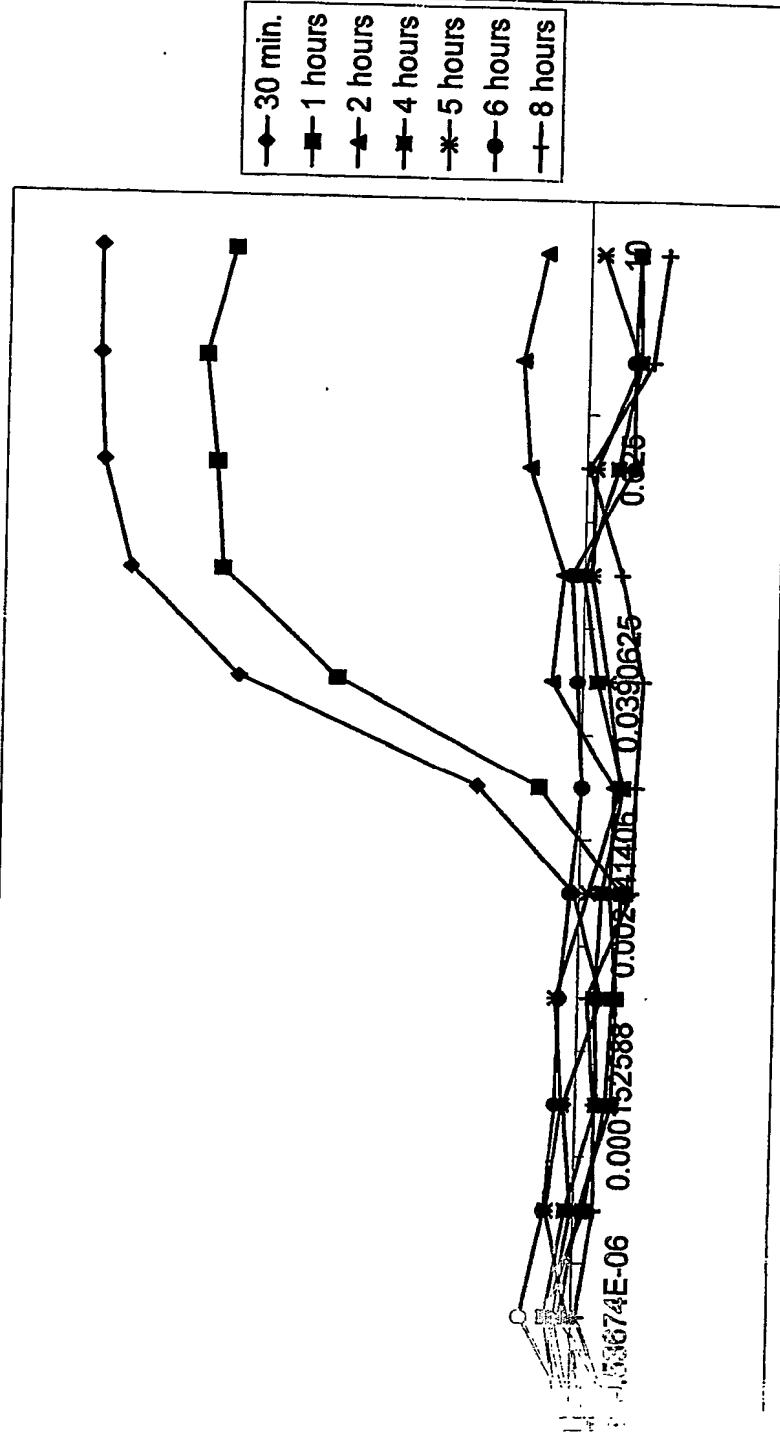
# Anti-viral-experiment with TOA protocol: 23°C



Concentration of T-20 (μM)

Figure 9A

# Anti-viral-experiment with TOA protocol: 37°C



Concentration of T-20 (µM)

Figure 9B



**This Page is Inserted by IFW Indexing and Scanning  
Operations and is not part of the Official Record**

**BEST AVAILABLE IMAGES**

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☒ **BLACK BORDERS**
- ☐ **IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- ☒ **FADED TEXT OR DRAWING**
- ☐ **BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- ☐ **SKEWED/SLANTED IMAGES**
- ☐ **COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- ☐ **GRAY SCALE DOCUMENTS**
- ☒ **LINES OR MARKS ON ORIGINAL DOCUMENT**
- ☐ **REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- ☐ **OTHER:** \_\_\_\_\_

**IMAGES ARE BEST AVAILABLE COPY.**

**As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.**